

FULL ARTICLE

High-resolution deep functional imaging of the whole mouse brain by photoacoustic computed tomography *in vivo*

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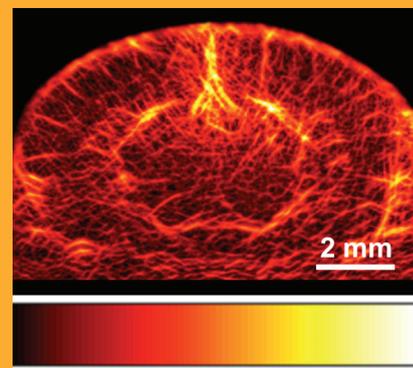
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Photoacoustic computed tomography (PACT) is a non-invasive imaging technique offering high contrast, high resolution, and deep penetration in biological tissues. We report a PACT system equipped with a high frequency linear transducer array for mapping the microvascular network of a whole mouse brain with the skull intact and studying its hemodynamic activities. The linear array was scanned in the coronal plane to collect data from different angles, and full-view images were synthesized from the limited-view images in which vessels were only partially revealed. We investigated spontaneous neural activities in the deep brain by monitoring the concentration of hemoglobin in the blood vessels and observed strong interhemispherical correlations between several chosen functional regions, both in the cortical layer and in the deep regions. We also studied neural activities during an epileptic seizure and ob-

served the epileptic wave spreading around the injection site and the wave propagating in the opposite hemisphere.



1. Introduction

In recent years, advanced imaging methodologies with increased spatial and temporal resolution have been used to extend our understanding of brain functions. Brain positron emission tomography (PET) is a metabolic imaging method that detects

the emissions from radioactive tracers that are injected into the blood stream and eventually reach the brain [1]. Functional magnetic resonance imaging (fMRI) measures brain activity by using the blood oxygenation level-dependent (BOLD) effect [2]. Both methods have drawbacks: PET uses ionizing radiation and has poor spatial resolution, and

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fMRI is too slow for studying fast hemodynamics. Recently, functional ultrasound imaging has successfully been used to monitor cerebral microvascular hemodynamics deep in the rat brain with high spatiotemporal resolution [3,4]. Optical imaging techniques, such as optical coherence tomography and multiphoton microscopy, have also been demonstrated for imaging neural activity with cellular and subcellular resolution [5,6], but the shallow penetration limits their observation to only the cortical layer (~1–2 mm).

Photoacoustic tomography (PAT) is a modality that detects ultrasonic waves generated by optical absorption of soft tissues under pulsed light illumination [7,8]. PAT is a nonionizing imaging technique, providing strong optical contrast and high ultrasonic spatial resolution at depths. By using multiple wavelength lasers to excite photoacoustic waves, PAT is able to simultaneously measure the concentrations of various chromophores in biological tissues or organs with distinct optical absorption properties. In particular, PAT has been successfully used to quantify the concentration and the oxygen saturation (sO_2) of hemoglobin, which are related to angiogenesis and hypermetabolism [7]. Substantially accurate sO_2 quantification has also been demonstrated in deep tissues [9]. By scanning a tightly focused laser beam, a PAT system can provide optical resolution photoacoustic microscopy (OR-PAM) [10,11]. Recently, OR-PAM has been used for high-speed, high-resolution mapping of the blood network and for studying the hemodynamics in the mouse brain [12]. Because OR-PAM relies on ballistic photons to achieve high resolution, its penetration depth in tissue is generally limited to ~1–2 mm. Photoacoustic computed tomography (PACT) detects ultrasonic waves generated by diffused photons and retrieves the absorption distribution through an inverse algorithm, allowing an imaging depth of up to several centimeters [13]. In this work, we used PACT to image the blood vascular network of a whole mouse brain through the intact skull, with high resolution and substantial depth. We then used the PACT system to study the hemodynamics of the mouse brain in the resting state and during an epileptic seizure. Our experiments show that functional PACT is a promising tool for studying neural activities in the deep brain, and potentially for revealing the mechanism of some brain disorders.

2. Material and methods

Figure 1 shows the schematic setup of linear array PACT for mouse brain imaging. In this work, 6–8 week-old female mice (Swiss Webster, Invigo)

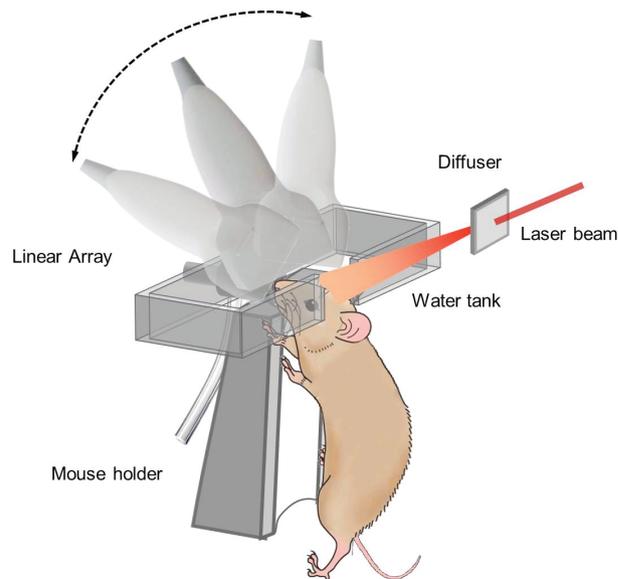


Figure 1 Schematic setup of linear array PACT for mouse brain imaging.

were used. Prior to imaging, the hair on the head of the mouse was removed by a depilatory cream. The mouse was then mounted onto a holder with its head fixed by a nose cone and a tooth bar. A rubber tube connecting the nose cone and an isoflurane vaporizer was used to deliver oxygen and anesthetic gas. The temperature of the mouse holder was regulated by a controller. The scalp of the mouse was removed, and ultrasound gel was applied before imaging. The animal procedures followed the laboratory animal protocols approved by the Animal Studies Committee of Washington University in St. Louis. The mouse was placed beneath the water tank, within an imaging window with dimensions of 8 cm × 10 cm. The imaging window was covered with plastic film at the bottom of the water tank, and the tank was filled with water for ultrasound coupling. A 5 cm wide window in one side of the tank wall allowed laser beam access to the mouse head. The laser beam used for exciting ultrasonic waves was provided by a Nd:YAG laser (Quantel, Brilliant B, 4–6 ns pulse duration, 10 Hz repetition rate). The laser beam was expanded and homogenized by an engineered diffuser (EDC-5-A-2 s, RPC Photonics, Inc.), resulting in a 2 cm-diameter illumination area on the mouse head. With a pulse energy of 200 mJ, the optical fluence on the head surface was 64 mJ/cm², which is below the American National Standards Institute (ANSI) safety limit of 100 mJ/cm² at 1064 nm. The excited photoacoustic (PA) waves were detected by a focused linear transducer array (LZ250, VisualSonics) consisting of 256 elements.

The linear array had a one-way bandwidth of 11 MHz around the central frequency of 21 MHz, providing a lateral resolution of 75 microns. The numerical aperture of the transducer array is 0.1, yielding a sectioning thickness of about 0.5 mm within the depth of focus, which covers the whole mouse brain. The detected signals were first amplified by homemade pre-amplifiers (256 channels, 26 dB Gain per channel) and then digitalized by a customized 256-channel data acquisition system (National Instruments, Customized PXI system, 14 bits) at a sampling rate of 250 MHz. The features within the imaging plane were reconstructed using the filtered universal back-projection algorithm [14].

3. Experimental results

To map the vessels in the coronal plane, the mouse was mounted with its cortical plane horizontal to the bottom of the water tank, and the linear array was oriented in the coronal plane, as shown in Figure 1. The surface of the linear array was about 1 cm away from the top of the head, and the diffused laser beam was obliquely incident on the head. The pulsed laser at 1064 nm was used to excite the PA waves, which came predominantly from oxy-

hemoglobin, a dominant chromophore in the blood stream. Due to the limited view of the linear array, only vessels normal to the acoustic axis could be reconstructed, but the vessels with their orientation deviated from the axis were almost invisible [15]. Figure 2(a–e) show reconstructed images of a typical coronal plane recorded from several representative view angles, each averaged over 200 acquisitions. To solve this limited-view problem, the linear array was mounted on a rotary stage (not shown in Figure 1) with its rotational axis centered at the mouse brain. The linear array was then scanned around the brain without changing the laser illumination to record images at view angles ranging from -76 to 76 degrees, with a step size of 4 degrees. The reconstructed bipolar images were converted into unipolar images by Hilbert transformation along each view's acoustic axis [16]. The processed images were then rotated back to recover the vascular distribution in the global coordinate system in which the brain was fixed, and the rotated images were summed to form a full-view unipolar image, as shown in Figure 2(f). The full-view image demonstrated that the PACT system equipped with the high frequency linear array can provide high resolution and deep penetration imaging of a whole mouse brain.

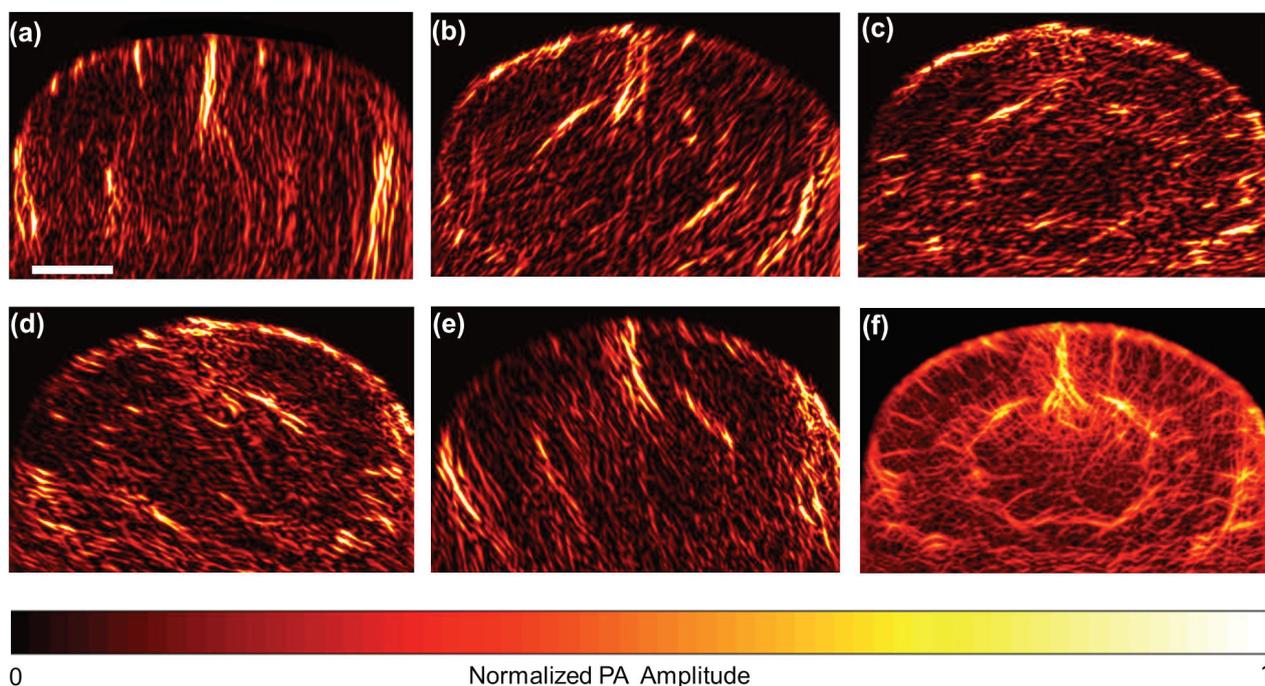


Figure 2 Coronal plane PACT imaging of a mouse brain through the intact skull (Bregma -1.0 mm). (a–e) Limited-view planar images reconstructed at view angles of -76 , -36 , 0 , 36 and 76 degrees, respectively. Each of these images was averaged over 200 acquisitions. (f) Full-view planar image combined from the 39 limited-view images. The scale bar is 2 mm.

Since light at the chosen laser wavelength is weakly absorbed in biological tissues, and changes in the concentrations of hemoglobin and other absorbers during neural activities are generally small, the local optical fluence in the mouse brain is assumed to be temporally invariant, enabling the PACT system to study the hemodynamic activities in the deep brain. As a first demonstration, we used our linear-array PACT system to study the resting-state functional connectivity (RSFC) in the coronal plane of a mouse brain. RSFC measures the temporal correlation of the low frequency, spontaneous hemodynamic fluctuations among spatially separated but functionally related regions in the brain [17,18]. The alteration in the functional connectivity properties is usually an indicator of such brain disorders as depression, Alzheimer's disease, and schizophrenia [19–21]. To explore the functional connectivity of a mouse brain in resting state, the linear array was fixed with its axis normal to the cortical plane, and 6000 images were acquired at a frame rate of 10 Hz. This study again used 6–8 week-old female ND4 Swiss Webster mice (Invigo), anesthetized with 0.8 % (vol/vol) isoflurane at an air flow rate of 0.8 L/min, a dose does not alter the functional connectivity patterns [22]. The PA signals due to scattered photons from the transducer surface were used to correct the fluctuations in the total laser pulse energy. Every set of 10 images was averaged to improve the signal-to-noise (SNR) ratio, and the resulting 600 images over a 10-minute acquisition were analyzed with a seed-based approach [23,24]. Basically, the reconstructed images were first filtered by a 2-D Gaussian smoothing kernel with standard deviation of 6 pixels. Then a band pass filter with a window of 0.009–0.08 Hz was applied on the time series of the images to reduce the effects caused by heart beating and breathing [24]. Finally, a seed was chosen and the temporal correlations of its PA signal with that of each pixel in the image were calculated. A high coefficient in the correlation map indicates a region that is functionally related to the seed region. The correlation maps for representative regions in the mouse brain were superimposed on the synthesized anatomic image obtained as described above. As shown in Figure 3(a–d), strong interhemispherical correlations were observed in the chosen functional regions, including the somatosensory, hippocampal, thalamic and retrohippocampal regions. The high correlations in the contralateral hemisphere were mostly from the regions symmetrically opposed to the seeds. Figure 3(e) also shows the time series of the filtered PA signals of a pair of correlated regions marked in white and black circles in Figure 3(a), displaying strong correlation in their fluctuations.

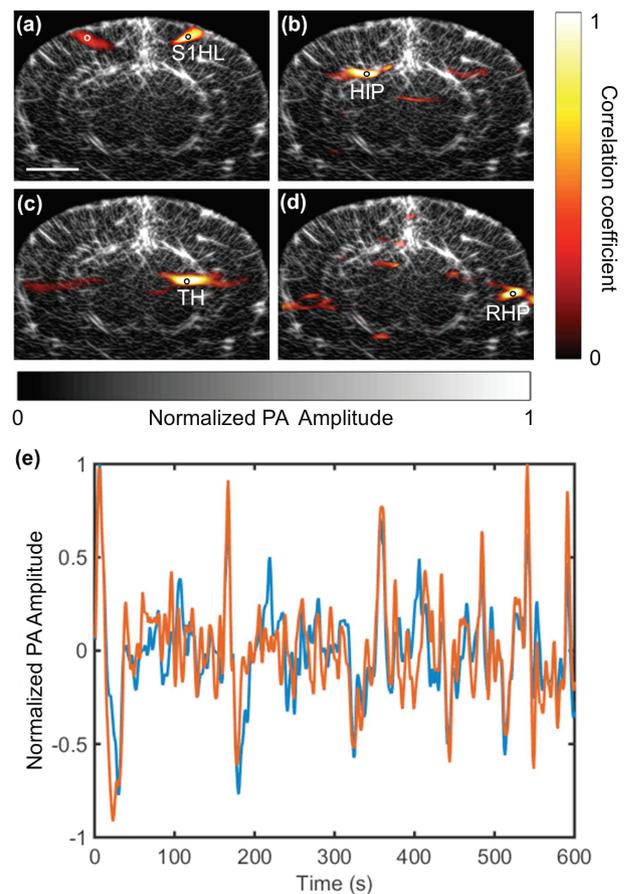


Figure 3 Functional connectivity in a mouse brain acquired by a linear-array PACT system. (a–d) The correlation maps calculated at four representative functional regions (black circles) are overlaid onto the anatomic image that was synthesized from multiple limited-view images (Bregma -1.0 mm). The scale bar is 2 mm. S1HL, somatosensory 1, hindlimb region; HIP, hippocampal region; TH, thalamus; RHP, retrohippocampal region. (e) Time series of the filtered PA signals of a pair of correlated regions as marked in (a). The red and blue curves are corresponding to the points marked in white and black circles, respectively. Amplitudes are normalized to the maximum.

PACT using a high frequency transducer array allows studying the hemodynamics in the mouse brain with high spatial and temporal resolutions. To demonstrate this capability, we used our linear-array PACT system to study epilepsy in a mouse model. An epileptic seizure in a mouse was induced by injection of 4-aminopyridine, a potassium channel blocker. A hole with a diameter of 1 mm was drilled in the skull prior to imaging, and 2 μ l of 4-aminopyridine solution at a concentration of 15 mM was injected into the brain through a 30 gauge needle. After the injection, the neural activities were imaged with the linear array fixed at an angle of 30 de-

grees. The images were acquired at a frame rate of 10 Hz, and every ten images were averaged into one, resulting in a temporal resolution of 1 s. The PA signal from the transducer surface due to scattered photons was used to correct the fluctuations in the total laser pulse energy. The fractional changes in the PA amplitude at different times after the injection are shown in Figure 4. As indicated, an increase in the PA amplitude was initially observed at the injection site and then spread out. Even more interesting, epileptic wave propagation was also observed in the opposite hemisphere (supplementary movie).

4. Conclusion

In summary, we have built a linear-array PACT system for imaging a whole mouse brain through the intact skull and for studying hemodynamic activities with high spatial resolution and deep penetration. The linear array was scanned in the coronal plane to collect images at different angles. Combing the lim-

ited-view images into a full-view image revealed the microvascular network in the brain. We investigated spontaneous neural activities in the deep brain by monitoring the concentration of hemoglobin in the blood vessels and observed strong interhemispherical correlations between chosen functional regions, both in the cortex and in the deep region. We also used linear-array PACT to study epilepsy in the mouse brain induced by an injection of a potassium channel blocker. We observed the epileptic wave spreading around the injection site and a corresponding wave propagating in the opposite hemisphere.

The imaging frame rate of our functional PACT system is lower than that of functional ultrasound imaging [3,4], but this limitation can be circumvented by employing a higher repetition-rate laser for photoacoustic excitation. However, PACT utilizes a different contrast mechanism from ultrasound imaging. Ultrasound tomography images the mechanical properties of soft tissue, and it has poor extravascular molecular contrasts. PACT measures the optical absorption of biomolecules, implementing

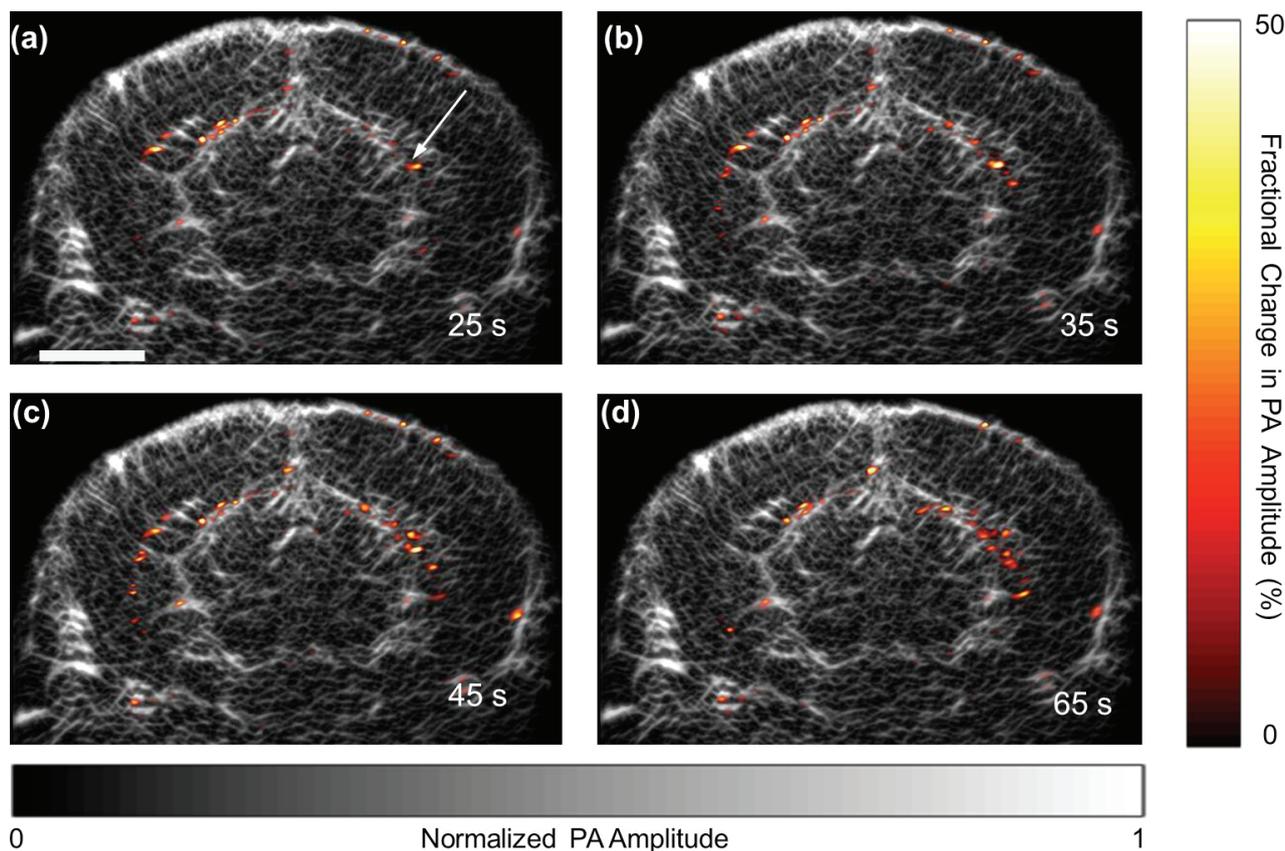


Figure 4 Epileptiform activities of a mouse brain during a seizure, as imaged by PACT. The fractional changes in the PA amplitude at different times after the injection of 4-aminopyridine solution are superimposed on the anatomic image (Bregma -1.0 mm). The scale bar is 2 mm, and the arrow indicates the injection site.

molecular imaging by manipulating the excitation wavelengths so that photoacoustic signals from the targeted molecules contribute mostly to the constructed images [25,26], and thus provide higher image contrast. Moreover, the oxygen saturation (sO₂) of hemoglobin, which cannot be obtained by ultrasound imaging but is closely related to brain activities, can be measured by PACT [7,9] when multiple wavelengths are used.

Supporting Information

Additional supporting information may be found in the online version of this article at the publisher's website.

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References

- [1] L. L. Firestone, F. Gyulai, M. Mintun, L. J. Adler, K. Urso, and P. M. Winter, *Anesth Analg* **82**(6), 1247–1251 (1996).
- [2] S. G. Kim, and S. Ogawa, *J Cereb Blood Flow Metab* **32**(7), 1188–1206 (2012).
- [3] E. Macé, G. Montaldo, I. Cohen, M. Baulac, M. Fink, and M. Tanter, *Nat Methods* **8**(8), 662–664 (2011).
- [4] B. F. Osmanski, S. Pezet, A. Ricobaraza, Z. Lenkei, and M. Tanter, *Nat Commun* **5** (2014).
- [5] Y. Chen, A. D. Aguirre, L. Ruvinskaya, A. Devor, D. A. Boas, and J. G. Fujimoto, *J Neurosci Methods* **178**(1), 162–173 (2009).
- [6] C. Tischbirek, A. Birkner, H. Jia, B. Sakmann, and A. Konnerth, *Proc Natl Acad Sci USA* **112**(36), 11377–11382 (2015).
- [7] H. F. Zhang, K. Maslov, G. Stoica, and L. V. Wang, *Nat Biotechnol* **24**(7), 848–851 (2006).
- [8] L. V. Wang, *Nat Photonics* **3**(9), 503–509 (2009).
- [9] S. Tzoumas, A. Nunes, I. Olefir, S. Stangl, P. Symvoulidis, S. Glasl, C. Bayer, G. Multhoff, and V. Ntziachristos, *Nat Commun* **7**, 12121 (2016).
- [10] K. Maslov, G. Stoica, and L. V. Wang, *Opt Lett* **30**(6), 625–627 (2005).
- [11] K. Maslov, H. F. Zhang, S. Hu, and L. V. Wang, *Opt Lett* **33**(9), 929–931 (2008).
- [12] J. Yao, L. Wang, J. M. Yang, K. I. Maslov, T. T. Wong, L. Li, C. H. Huang, J. Zou, and L. V. Wang, *Nat Methods* **12**(5), 407–410 (2015).
- [13] L. V. Wang and S. Hu, *Science* **335**(6075), 1458–1462 (2012).
- [14] M. Xu and L. V. Wang, *Phys Rev E* **71**(1), 016706 (2005).
- [15] Y. Xu, L. V. Wang, G. Ambartsoumian, and P. Kuchment, *Med Phys* **31**(4), 724–733 (2004).
- [16] G. Li, L. Li, L. Zhu, J. Xia, and L. V. Wang, *J Biomed Opt* **20**(6), 066010 (2015).
- [17] B. Biswal, F. Zerrin Yetkin, V. M. Haughton, and J. S. Hyde, *Magn Reson Med* **34**(4), 537–541 (1995).
- [18] M. D. Fox and M. E. Raichle, *Nat Rev Neurosci* **8**(9), 700–711 (2007).
- [19] M. D. Greicius, B. H. Flores, V. Menon, G. H. Glover, H. B. Solvason, H. Kenna, A. L. Reiss, and A. F. Schatzberg, *Biol Psychiatry* **62**(5), 429–437 (2007).
- [20] R. L. Buckner, J. Sepulcre, T. Talukdar, F. M. Krienen, H. Liu, T. Hedden, J. R. Andrews-Hanna, R. A. Sperling, and K. A. Johnson, *J Neurosci* **29**(6), 1860–1873 (2009).
- [21] S. M. Lawrie, C. Buechel, H. C. Whalley, C. D. Frith, K. J. Friston, and E. C. Johnstone, *Biol Psychiatry* **51** (12), 1008–1011 (2002).
- [22] J. Grandjean, A. Schroeter, I. Batata, and M. Rudin, *Neuroimage* **102**, 838–847 (2014).
- [23] M. D. Fox, D. Zhang, A. Z. Snyder, and M. E. Raichle, *J Neurophysiol* **101**(6), 3270–3283 (2009).
- [24] M. Nasiravanaki, J. Xia, H. Wan, A. Q. Bauer, J. P. Culver, and L. V. Wang, *Proc Natl Acad Sci USA* **111** (1), 21–26 (2014).
- [25] J. Yao, A. A. Kaberniuk, L. Li, D. M. Shcherbakova, R. Zhang, L. Wang, G. Li, V. V. Verkhusha, and L. V. Wang, *Nat Methods* **13**(1), 67–73 (2016).
- [26] A. P. Jathoul, J. Laufer, O. Ogunlade, B. Treeby, B. Cox, E. Zhang, P. Johnson, A. R. Pizzey, B. Philip, T. Marafiotti, and M. F. Lythgoe, *Nat Photonics* **9**(4), 239–246 (2015).